

# **Mouse tPA Total Antigen ELISA**

Product Number: UP58

Store at 4°C

FOR RESEARCH USE ONLY Document Control Number: UP58.220617

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# Enzyme Immunoassay for Mouse Tissue Plasminogen Activator (tPA) Total Antigen

For Research Use Only

## INTRODUCTION

The Mouse tPA total antigen assay is intended for the quantitative determination of total plasminogen activator antigen in biological fluids.

Tissue plasminogen activator (tPA) is a serine protease that converts plasminogen to the active serine protease plasmin in the blood fibrinolytic system<sup>1,2</sup>. It also plays an important role in the removal of incipient thrombi<sup>3</sup>. tPA is widely used for the thrombolytic treatment of acute myocardial infarction<sup>3</sup>.

## PRINCIPLES OF PROCEDURE

Mouse tPA will bind to the capture antibody coated on the microtiter plate. Free and complexed enzyme will react with the capture antibody on the plate. A standard calibration curve is prepared using dilutions of tPA along with the samples to be measured. After appropriate washing steps, monoclonal anti-mouse tPA primary antibody binds to the captured enzyme. Excess antibody is washed away and bound monoclonal antibody is then reacted with the secondary antibody conjugated to horseradish peroxidase. TMB substrate is used for color development at 450 nm.

## MATERIALS PROVIDED

Component	Component Contents			Cat. No.
Coated Plate	Capture antibody coated 96-well plate	1 plate	4°C	UP58a
Standard	Mouse tPA standard	1 vial	4°C	UP58b
Primary Antibody	Monoclonal anti-mouse tPA antibody	1 vial	4°C	UP58c
Secondary Antibody	Anti-mouse HRP conjugated antibody	1 vial	4°C	UP58d
Wash Buffer	10x solution for washing plate	50 mL	4°C	UP58e
Substrate	TMB substrate	10 mL	4°C	UP58f

# MATERIALS NEEDED BUT NOT PROVIDED

- 1. Pipettes covering 0-10  $\mu$ l and 200-1000  $\mu$ l and tips
- 2. 12-channel pipette covering 30-300 µl
- 3. 1N H<sub>2</sub>SO<sub>4</sub>
- 4. DI water
- 5. Microtiter plate spectrophotometer with a 450 nm filter
- 6. Microtiter plate shaker with uniform horizontally circular movement up to 300 rpm

# STORAGE CONDITIONS

- 1. Store this kit and its components at 4°C until use.
- 2. Reconstituted standards and primary may be stored at -70°C for later use. **DO NOT** freeze/thaw the Standards and Primary Antibody more than once.

#### PROCEDURAL NOTES

- 1. Use aseptic technique when opening and dispensing reagents.
- 2. This kit is designed to work properly as provided and instructed. Additions, deletions, or substitutions to the procedure or reagents are not recommended, as they may be detrimental to the assay.
- 3. Exercise universal precautions during the performance or handling of this kit or any component contained therein.

## SAMPLE COLLECTION AND PREPARATION

Samples of plasma, serum, cell culture media or other biological fluids may be applied directly to the plate.

The assay measures tPA antigen in the 0.1-50 ng/ml range. Samples giving tPA levels above 50 ng/ml should be diluted in plasma devoid of tPA or 3% BSA Blocking Buffer.

## REAGENT PREPARATION

- 1. **10x Wash Buffer:** Dilute to 1x by adding 450 mL of DI water to the 50 mL of 10x Wash Buffer prior to use.
- 2. **TBS Buffer (100 mL):** 0.1 M Tris-HCL, 0.15 M NaCl, pH 7.4
- 3. 3% BSA Blocking Buffer (20 mL): 3% BSA (w/v) in TBS Buffer
- 4. **Standard:** Reconstitute with 1mL of the 3% BSA blocking buffer to obtain a 1000 ng/mL standard stock solution and vortex gently to mix. Prepare immediately prior to use.
- 5. **Primary Antibody:** Reconstitute with 10mL of 3% BSA Blocking Buffer and vortex gently to mix. Prepare immediately prior to use.
- 6. **Secondary Antibody:** Dilute 1uL with 10mL of 3% BSA Blocking Buffer and vortex gently to mix. Prepare immediately prior to use.

# STANDARD PREPARATION

Reconstitute the Standard as directed on the vial to give a 100 ng/mL Standard Stock Solution. **Do not prepare the standards until you are ready to apply them to the plate.** 

**Table 1: Preparation of Standard Curve** 

Standard	tPA Concentration (ng/mL)	Blocking Buffer (µL)	Transfer Volume (µL)	Transfer Source	Final Volume (µL)	
$S_9$	50	950	50	Stock Vial	500	
$S_8$	25	500	500	S <sub>9</sub>	600	
$S_7$	10	600	400	$S_8$	500	
$S_6$	5	500	500	S <sub>7</sub>	600	
$S_5$	2	600	400	$S_6$	500	
$S_4$	1	500	500	$S_5$	500	
$S_3$	0.5	500	500	S <sub>4</sub>	500	
$S_2$	0.25	500	500	$S_3$	600	
$S_1$	0.1	600	400	$S_2$	1,000	
$B_0$	0	500			500	

## **ASSAY PROCEDURE**

- 1. Add  $100 \mu l$  of the Standards and unknowns to the wells in duplicate. Shake the plate at 300 rpm for 30 minutes at room temperature (RT). See Scheme 1 below for a sample plate layout.
- 2. Wash the plate three times with 300  $\mu$ L of Wash Buffer. Remove excess Wash Buffer by gently tapping the plate on a paper towel.
- 3. Add 100 µl of the Primary Antibody to each well. Shake the plate at 300 rpm for 30 minutes at RT.
- 4. Wash the plate three times with 300  $\mu$ L of Wash Buffer as in step 2.
- 5. Add 100 µl of the Primary Antibody to each well. Shake the plate at 300 rpm for 30 minutes at RT.
- 6. Wash the plate three times with 300  $\mu$ L of Wash Buffer as in step 2.
- 7. Add 100 µl of TMB Substrate to each well. Shake the plate at 300 rpm for 2-10 minutes at RT.
- 8. Stop the reaction by adding 50  $\mu$ l of 1N H<sub>2</sub>SO<sub>4</sub> and read the plate at 450 nm.

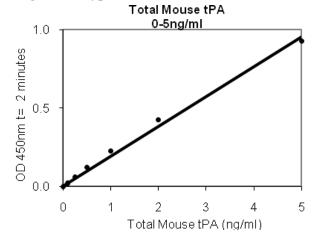
Scheme 1:	Sample	<b>Plate</b>	Layout
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	1	2	3	4	5	6	7	8	9	10	11	12
A	S9	S <sub>8</sub>	S7	S <sub>6</sub>	S <sub>5</sub>	S4	S <sub>3</sub>	S <sub>2</sub>	S <sub>1</sub>	В0	$U_1$	U <sub>2</sub>
В	S9	S <sub>8</sub>	<b>S</b> 7	S <sub>6</sub>	S <sub>5</sub>	S4	S3	S <sub>2</sub>	$s_1$	B <sub>0</sub>	$U_1$	$U_2$
C	U3	U4	U5	U6	U7	U8	U9	$U_{10}$	U11	U12	U13	$U_{14}$
D	U3	U4	U5	$U_6$	U7	$U_8$	U9	$U_{10}$	$U_{11}$	U12	U13	$U_{14}$
								$U_{22}$				
F	$U_{15}$	$U_{16}$	$U_{17}$	$U_{18}$	U19	$U_{20}$	$U_{21}$	$U_{22}$	$U_{23}$	$U_{24}$	U25	$U_{26}$
								U34				
Н	U27	U28	U29	U30	U31	U32	U33	U34	U35	U36	U37	U38

## **CALCULATIONS**

- 1. Average all the duplicate wells and subtract the zero point standard (B<sub>0</sub>) from all standards and unknowns to obtain the corrected absorbance.
- 2. Plot the A<sub>450</sub> against the concentration of tPA in the standards.
- 3. Fit a straight line through the points using a linear fit procedure.
- 4. Calculate the uPA concentrations in the unknowns using the standard curve. See Figure 1 bellow for an example of a typical standard curve.

Figure 1: Typical Standard Curve



#### EXPECTED VALUES

The concentration level of tPA antigen in mouse plasma has been reported to be 2.5+/-1.0 ng/ml<sup>4</sup>. In house testing of pooled normal mouse plasma in citrate indicates tPA levels vary by mouse strain:

Strain	Active tPA	Total tPA
NSA/CF-1	9.9 ng/mL	9.4 ng/mL
C57BL6	1.4 ng/mL	2.4 ng/mL
CD-1	0.4  ng/mL	0.4  ng/mL

Abnormalities in tPA levels have been reported in the following conditions:

- Venous Thrombosis: Locally applied tPA reduces thrombus formation after vascular injury<sup>9</sup>.
- Ischemic Diseases: tPA may affect the course of ischemic diseases<sup>5</sup>.
- Pathological Infarction: tPA may prevent or limit pathological infarction and improve neurological functions<sup>6</sup>. Usage of tPA at the onset of ischemic stroke improves clinical outcome<sup>7</sup>.
- Blood-Brain Barrier: is necessary and sufficient to directly increase the vascular permeability in the early stages of BBB opening8.

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