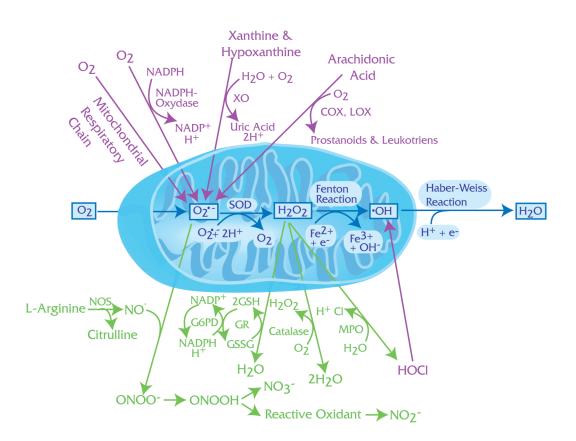


Antioxidant Assays: How Do They Compare

Denis Callewaert, Ph.D. President, Oxford Biomedical Research, Inc. Emeritus Professor of Chemistry, Oakland University While molecular oxygen is essential for a wide array of organisms, O₂ and reactive oxygen species (ROS) can be toxic. Therefore, humans and other organisms that utilize oxygen must have very effective antioxidant systems. Indeed, the antioxidant capabilities of higher organisms are due to the sum of enzymatic and non-enzymatic mechanisms. These include hydrophilic (uric acid, vitamin C, bilirubin, glutathione) as well as hydrophobic (most importantly vitamin E) antioxidants as well as several enzymes that either directly metabolize ROS (e.g. superoxide dismutase and catalase), or replenish the supply of reduced ROS scavengers (e.g. glutathione reductase).



Antioxidant capacity is influenced by a wide range of factors. It is therefore important to be able to quantitatively assess the total antioxidant power or capacity within biological specimens. Over the years, a number of assays purporting to measure total antioxidant power have been reported. However, most of these have significant limitations [1,2]. In 2003, a new method simply called the Total Antioxidant Power (TAP) assay was patented. This method, which is available exclusively through Oxford Biomedical Research, overcomes most of the problems that have been identified with other antioxidant assays and has been widely adopted for *in vitro* and *in vivo* studies in many laboratories. First, it is appropriate to briefly summarize the pros and cons of the antioxidant assays that were developed previously.

Common Methods for Measuring Antioxidants

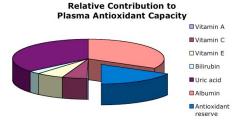
TEAC: The Trolox Antioxidant Capacity assay [3] can be used to measure antioxidant capacity of hydrophilic substances, but suffers from interference by peroxidases and nonlinear results upon sample dilution.

TRAP: The Total Radical Trapping Antioxidant Parameter assay [4] was modified in 1995 to correct for interference due to plasma proteins or sample dilution [5]. While it is fast and convenient, it is not clear to what extent lipid-soluble antioxidants contribute to the values obtained and does not measure the ability of plasma to interfere with lipid peroxidation due to the azo compound (ABAP) used in the assay.

FRAP: The Ferric Reducing Ability of Plasma assay [6] is responsive to uric acid but does not adequately measure the antioxidant activity of many important antioxidants such as ascorbic acid, glutathione and albumin [6,7]. The assay further suffers from low precision [1].

ORAC: The Oxygen Radical Absorbance Capacity assay [8,9] is a simple, and reliable way to measure the peroxyl radical absorbing capacity of serum or other biological fluids. The meaning of the ORAC index is less clear [2]. For the most potent ROS it provides results comparable to TRAP. However, for less reactive ROS the results obtained correlate better with other methods.

TAP: The Total Antioxidant Power assay [10] quantifies the combined action of all antioxidants present in the sample capable of reduction of Cu2+ followed by formation of a stable Cu+-bathocuproine complex that can be quantified at 480 nm. The redox potential for this reaction is ideal for the accurate determination of the combined antioxidant activity in a specimen that results from all of its constituents including vitamins, proteins, glutathione, uric acid, etc. Although introduced relatively recently, this assay has already been applied to a wide range of biological systems [11-13] and is proving particularly valuable for reproducible, reliable and convenient for studies of human body fluids [14-19]. This assay provides very reliable results. However, certain pathological conditions may unduly influence the results. For example, in diabetics, uric acid exhibits pro-oxidant activity. In these patients, pre-incubation with uricase allows for quantification of the true antioxidant power. Thus, used appropriately, the TAP assay can be used to reliably quantify the total antioxidant activity of a wide range of specimens. These include physiological fluids as well as food and beverage products.



Oxford Biomedical Research Page 3 of 4

Oxfordbiomed.com

References:

- McAnalley S, Koepke CM, Le L, Vennum E, McAlley B, (2003) In Vitro methods for testing antioxidant 1 potential: a review. Glycoscience & Nutrition 4:1-9.
- 2. Perez DD, Leighton F, Aspee A, Aliaga C and Lissi E (2000) A comparison of methods employed to evaluate antioxidant capabilities. Biol. Res. 33 (2).
- NJ Miller, C Rice-Evans, MJ Davies, V Gopinathan, and A Milner (1993) A novel method for measuring 3. antioxidant capacity and its application to monitoring the antioxidant status in premature neonates. Clin Sci (Lond), 84: 407-12.
- 4. DD Wayner, GW Burton, KU Ingold, and S Locke (1985) Quantitative measurement of the total, peroxyl radical-trapping antioxidant capability of human blood plasma by controlled peroxidation. The important contribution made by plasma proteins. FEBS Lett, 187: 33-7.
- 5. A Ghiselli, M Serafini, G Maiani, E Azzini, and A Ferro-Luzzi (1995) A fluorescence-based method for measuring total plasma antioxidant capability. Free Radic Biol Med, 18: 29-36.
- 6. IF Benzie and JJ Strain (1996) The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay. Anal Biochem, 239: 70-6.
- SB Lotito and B Frei (2004) The increase in human plasma antioxidant capacity after apple consumption is 7. due to the metabolic effect of fructose on urate, not apple-derived antioxidant flavonoids. Free Radic Biol Med, 37: 251-8.
- G Cao, HM Alessio, and RG Cutler (1993) Oxygen-radical absorbance capacity assay for antioxidants. Free 8 Radic Biol Med, 14: 303-11.
- 9. B Ou, M Hampsch-Woodill, and RL Prior (2001) Development and validation of an improved oxygen radical absorbance capacity assay using fluorescein as the fluorescent probe. J Agric Food Chem, 49: 4619-26.
- 10. DaCruz G (2003) Use of bathocuproine for the evaluation of the antioxidant power in liquids and solutions U.S. Patent 6.613.577
- 11. Nicholas J. Hudson, Sigrid A. Lehnert, Aaron B. Ingham, Beth Symonds, Craig E. Franklin, and Gregory S. Harper (2006) Lessons from an estivating frog: sparing muscle protein despite starvation and disuse. Am J Physiol Regulatory Integrative Comp Physiol, 290: R836 - R843.
- 12. Anthony Fardet, Cécile Canlet, Gaëlle Gottardi, Bernard Lyan, Rafaël Llorach, Christian Rémésy, André Mazur, Alain Paris, and Augustin Scalbert (2007) Whole-Grain and Refined Wheat Flours Show Distinct Metabolic Profiles in Rats as Assessed by a ¹H NMR-Based Metabonomic Approach. J. Nutr., 137: 923 -929.
- 13. Atsunori Nakao, Kei Kimizuka, Donna B. Stolz, Joao Seda Neto, Takashi Kaizu, Augustine M. K. Choi, Takashi Uchiyama, Brian S. Zuckerbraun, Michael A. Nalesnik, Leo E. Otterbein, and Noriko Murase (2003) Carbon Monoxide Inhalation Protects Rat Intestinal Grafts from Ischemia/Reperfusion Injury. Am. J. Pathol., 163: 1587.
- 14. deMartino M, Chiarelli F, Moriondo M, Torello M, Azzari C and Galli L, (2001) Restored antioxidant capacity parallels the immunologic and virologic improvement in children with perinatal human immunodeficiency virus infection receiving highly active antiretroviral therapy.lin Immunol. 100: 82-86 (2001).
- 15. K. Matsuno, M. Osako, and M. Usui (2005) Total Antioxidant Power in the Vitreous of Diabetic Retinopathy Patients. Invest. Ophthalmol. Vis. Sci., 46: 381.
- 16. Theresa O Scholl, Maria Leskiw, Xinhua Chen, Melissa Sims, and T Peter Stein (2005) Oxidative stress, diet, and the etiology of preeclampsia. Am. J. Clinical Nutrition, 81: 1390 - 1396.
- 17. Atsunori Nakao, Noriko Murase, Chien Ho, Hideyoshi Toyokawa, Timothy R. Billiar, and Shinichi Kanno (2005) Biliverdin Administration Prevents the Formation of Intimal Hyperplasia Induced by Vascular Injury. Circulation, 112: 587 - 591.
- 18. Lihong Jia, Zhongbo Liu, Lijuan Sun, Sheldon S. Miller, Bruce N. Ames, Carl W. Cotman, and Jiankang Liu (2007) Acrolein, a Toxicant in Cigarette Smoke, Causes Oxidative Damage and Mitochondrial Dysfunction in RPE Cells: Protection by (R)--Lipoic Acid Invest. Ophthalmol. Vis. Sci., 48: 339 - 348.
- 19. Futoshi Okada, Masanobu Kobayashi, Hiroki Tanaka, Tokushige Kobayashi, Hiroshi Tazawa, Yoshihito luchi, Kunishige Onuma, Masuo Hosokawa, Mary C. Dinauer, and Nicholas H. Hunt (2006) The Role of Nicotinamide Adenine Dinucleotide Phosphate Oxidase-Derived Reactive Oxygen Species in the Acquisition of Metastatic Ability of Tumor Cells. Am. J. Pathol. 169: 294 - 302.

1-800-692-4633