

Enzyme Immunoassay for Mouse Tissue Plasminogen Activator (tPA) Total Antigen

For Research Use Only

INTRODUCTION

The Mouse tPA total antigen assay is intended for the quantitative determination of total plasminogen activator antigen in biological fluids.

Tissue plasminogen activator (tPA) is a serine protease that converts plasminogen to the active serine protease plasmin in the blood fibrinolytic system^{1,2}. It also plays an important role in the removal of incipient thrombi³. tPA is widely used for the thrombolytic treatment of acute myocardial infarction³.

PRINCIPLES OF PROCEDURE

Mouse tPA will bind to the capture antibody coated on the microtiter plate. Free and complexed enzyme will react with the capture antibody on the plate. A standard calibration curve is prepared using dilutions of tPA along with the samples to be measured. After appropriate washing steps, monoclonal anti-mouse tPA primary antibody binds to the captured enzyme. Excess antibody is washed away and bound monoclonal antibody is then reacted with the secondary antibody conjugated to horseradish peroxidase. TMB substrate is used for color development at 450 nm.

MATERIALS PROVIDED

| Component | Contents | Quantity | Storage | Cat. No. |
|--------------------|---------------------------------------|----------|---------|----------|
| Coated Plate | Capture antibody coated 96-well plate | 1 plate | 4°C | UP58a |
| Standard | Mouse tPA standard | 1 vial | 4°C | UP58b |
| Primary Antibody | Monoclonal anti-mouse tPA antibody | 1 vial | 4°C | UP58c |
| Secondary Antibody | Anti-mouse HRP conjugated antibody | 1 vial | 4°C | UP58d |
| Wash Buffer | 10x solution for washing plate | 50 mL | 4°C | UP58e |
| Substrate | TMB substrate | 10 mL | 4°C | UP58f |

MATERIALS NEEDED BUT NOT PROVIDED

1. Pipettes covering 0-10 μ l and 200-1000 μ l and tips
2. 12-channel pipette covering 30-300 μ l
3. 1N H₂SO₄
4. DI water
5. Microtiter plate spectrophotometer with a 450 nm filter
6. Microtiter plate shaker with uniform horizontally circular movement up to 300 rpm

STORAGE CONDITIONS

1. Store this kit and its components at 4°C until use.
2. Reconstituted standards and primary may be stored at -70°C for later use. **DO NOT** freeze/thaw the Standards and Primary Antibody more than once.

PROCEDURAL NOTES

1. Use aseptic technique when opening and dispensing reagents.
2. This kit is designed to work properly as provided and instructed. Additions, deletions, or substitutions to the procedure or reagents are not recommended, as they may be detrimental to the assay.
3. Exercise universal precautions during the performance or handling of this kit or any component contained therein.

SAMPLE COLLECTION AND PREPARATION

Samples of plasma, serum, cell culture media or other biological fluids may be applied directly to the plate.

The assay measures tPA antigen in the 0.1-50 ng/ml range. Samples giving tPA levels above 50 ng/ml should be diluted in plasma devoid of tPA or 3% BSA Blocking Buffer.

REAGENT PREPARATION

1. **10x Wash Buffer:** Dilute to 1x by adding 450 mL of DI water to the 50 mL of 10x Wash Buffer prior to use.
2. **TBS Buffer (100 mL):** 0.1 M Tris-HCL, 0.15 M NaCl, pH 7.4
3. **3% BSA Blocking Buffer (20 mL):** 3% BSA (w/v) in TBS Buffer
4. **Standard:** Reconstitute as directed on the vial to obtain a 1000 ng/mL standard stock solution and vortex gently to mix. Prepare immediately prior to use.
5. **Primary Antibody:** Reconstitute with 3% BSA Blocking Buffer as directed on the vial and vortex gently to mix. Prepare immediately prior to use.
6. **Secondary Antibody:** Dilute with 3% BSA Blocking Buffer as directed on the vial and vortex gently to mix. Prepare immediately prior to use.

STANDARD PREPARATION

Reconstitute the Standard as directed on the vial to give a 100 ng/mL Standard Stock Solution. **Do not prepare the standards until you are ready to apply them to the plate.**

Table 1: Preparation of Standard Curve

| Standard | tPA Concentration (ng/mL) | Blocking Buffer (µL) | Transfer Volume (µL) | Transfer Source | Final Volume (µL) |
|----------------|---------------------------|----------------------|----------------------|-----------------|-------------------|
| S ₉ | 50 | 950 | 50 | Stock Vial | 500 |
| S ₈ | 25 | 500 | 500 | S ₉ | 600 |
| S ₇ | 10 | 600 | 400 | S ₈ | 500 |
| S ₆ | 5 | 500 | 500 | S ₇ | 600 |
| S ₅ | 2 | 600 | 400 | S ₆ | 500 |
| S ₄ | 1 | 500 | 500 | S ₅ | 500 |
| S ₃ | 0.5 | 500 | 500 | S ₄ | 500 |
| S ₂ | 0.25 | 500 | 500 | S ₃ | 600 |
| S ₁ | 0.1 | 600 | 400 | S ₂ | 1,000 |
| B ₀ | 0 | 500 | --- | --- | 500 |

ASSAY PROCEDURE

1. Add 100 μ l of the Standards and unknowns to the wells in duplicate. Shake the plate at 300 rpm for 30 minutes at room temperature (RT). See Scheme 1 below for a sample plate layout.
2. Wash the plate three times with 300 μ L of Wash Buffer. Remove excess Wash Buffer by gently tapping the plate on a paper towel.
3. Add 100 μ l of the Primary Antibody to each well. Shake the plate at 300 rpm for 30 minutes at RT.
4. Wash the plate three times with 300 μ L of Wash Buffer as in step 2.
5. Add 100 μ l of the Primary Antibody to each well. Shake the plate at 300 rpm for 30 minutes at RT.
6. Wash the plate three times with 300 μ L of Wash Buffer as in step 2.
7. Add 100 μ l of TMB Substrate to each well. Shake the plate at 300 rpm for 2-10 minutes at RT.
8. Stop the reaction by adding 50 μ l of 1N H₂SO₄ and read the plate at 450 nm.

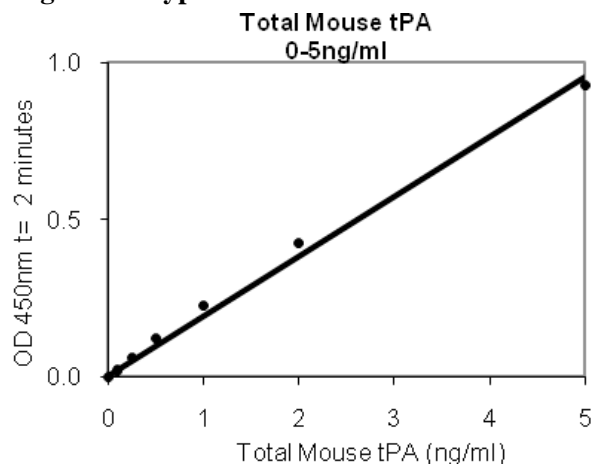
Scheme 1: Sample Plate Layout

| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|---|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| A | S9 | S8 | S7 | S6 | S5 | S4 | S3 | S2 | S1 | B0 | U1 | U2 |
| B | S9 | S8 | S7 | S6 | S5 | S4 | S3 | S2 | S1 | B0 | U1 | U2 |
| C | U3 | U4 | U5 | U6 | U7 | U8 | U9 | U10 | U11 | U12 | U13 | U14 |
| D | U3 | U4 | U5 | U6 | U7 | U8 | U9 | U10 | U11 | U12 | U13 | U14 |
| E | U15 | U16 | U17 | U18 | U19 | U20 | U21 | U22 | U23 | U24 | U25 | U26 |
| F | U15 | U16 | U17 | U18 | U19 | U20 | U21 | U22 | U23 | U24 | U25 | U26 |
| G | U27 | U28 | U29 | U30 | U31 | U32 | U33 | U34 | U35 | U36 | U37 | U38 |
| H | U27 | U28 | U29 | U30 | U31 | U32 | U33 | U34 | U35 | U36 | U37 | U38 |

CALCULATIONS

1. Average all the duplicate wells and subtract the zero point standard (B0) from all standards and unknowns to obtain the corrected absorbance.
2. Plot the A₄₅₀ against the concentration of tPA in the standards.
3. Fit a straight line through the points using a linear fit procedure.
4. Calculate the uPA concentrations in the unknowns using the standard curve. See Figure 1 below for an example of a typical standard curve.

Figure 1: Typical Standard Curve



EXPECTED VALUES

The concentration level of tPA antigen in mouse plasma has been reported to be 2.5+/-1.0 ng/ml⁴. In house testing of pooled normal mouse plasma in citrate indicates tPA levels vary by mouse strain:

| Strain | Active tPA | Total tPA |
|----------|------------|-----------|
| NSA/CF-1 | 9.9 ng/mL | 9.4 ng/mL |
| C57BL6 | 1.4 ng/mL | 2.4 ng/mL |
| CD-1 | 0.4 ng/mL | 0.4 ng/mL |

Abnormalities in tPA levels have been reported in the following conditions:

- Venous Thrombosis: Locally applied tPA reduces thrombus formation after vascular injury⁹.
- Ischemic Diseases: tPA may affect the course of ischemic diseases⁵.
- Pathological Infarction: tPA may prevent or limit pathological infarction and improve neurological functions⁶. Usage of tPA at the onset of ischemic stroke improves clinical outcome⁷.
- Blood-Brain Barrier: is necessary and sufficient to directly increase the vascular permeability in the early stages of BBB opening⁸.

REFERENCES

1. Quax, Paul H. A., *et al.*: (1990) J. Biol. Chem.; **265(26)**:15560-15563
2. Kristensen, Paul, *et al.*: (1985) J. Cell Biol.; **(101)**:305-311
3. Camani, Chantal, *et al.*: (1994) J. Biol. Chem.; **269(8)**:5770-5775
4. Declerck, P. J., *et al.*: (1995) Thromb Haemostas.; **74(5)**:1305-1309
5. Nassar, T., *et al.*: (2003) Blood; **103(3)**:897-902
6. Sakurama, T., *et al.*: (1994) Stroke; **25**:451-456
7. The National Institute of Neurological Disorders and Stroke rt-PA Stroke Study Group: (1995) N Engl J Med.; **333(24)**:1581-1588
8. Yepes, M., *et al.*: (2003) J Clin Invest.; **112(10)**:1483-1485
9. Underwood, M. J., *et al.*: (1993) Cardiovasc. Res.; **27(12)**:2270-2273

DISCLAIMER

This information is believed to be correct but does not purport to be all-inclusive and shall be used only as a guide. Oxford Biomedical Research, Inc. shall not be held liable for any damage resulting from handling or from contact with the above product. See catalog for additional terms and conditions of sale.

TECHNICAL SUPPORT

If you need technical information or assistance with assay procedures, please call our Technical Support Department at 800-692-4633 or 248-852-8815. Our staff will answer your questions about this or any other product in the Oxford Biomedical line.

GUARANTEE AND LIMITATION OF REMEDY

Oxford Biomedical Research, Inc. makes no guarantee of any kind, expressed or implied, which extends beyond the description of the material in this ELISA kit, except that these materials and this kit will meet our specifications at the time of delivery. Buyer's remedy and Oxford Biomedical Research, Inc.'s sole liability hereunder is limited to, at Oxford Biomedical Research, Inc.'s option, refund of the purchase price of, or the replacement of, material that does not meet our specification. By acceptance of our products, Buyer indemnifies and holds Oxford Biomedical Research, Inc. harmless against, assumes all liability for the consequence of its use or misuse by the Buyer, its employees, or others. Said refund or replacement is conditioned of Buyer notifying Oxford Biomedical Research, Inc. within thirty (30) days of the receipt of product. Failure of Buyer to give said notice within said thirty (30) days shall constitute a waiver by the Buyer of all claims hereunder with respect to said material(s).

Oxford Biomedical Research, Inc.
P.O. Box 522
Oxford, MI 48371 U.S.A.

Orders: 800-692-4633
Technical Service: 248-852-8815
Fax: 248-852-4466
E-mail: info@oxfordbiomed.com

Made in the U.S.A.